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Adenovirus-mediated *in vivo* gene transfer and expression in normal rat liver

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Replication deficient, recombinant adenovirus (Ad) vectors do not require target cell replication for transfer and expression of exogenous genes and thus may be useful for *in vivo* gene therapy in hepatocytes. *In vitro*, primary cultures of rat hepatocytes infected with a recombinant Ad containing a human α 1-antitrypsin cDNA (Ad- α 1AT) synthesized and secreted human α 1AT for 4 weeks. In rats, *in vivo* intraportal administration of a recombinant Ad containing the *E. coli* lacZ gene, was followed by expression of β -galactosidase in hepatocytes 3 days after infection. Intraportal infusion of Ad- α 1AT produced detectable serum levels of human α 1AT for 4 weeks. Thus, targeted gene expression has been achieved in the liver, albeit at low levels, suggesting that adenovirus vectors may be a useful means for *in vivo* gene therapy in liver disorders.

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A significant challenge in the development of gene therapy for hereditary and acquired disorders in man is the expression of the therapeutic gene in the relevant cell population. The hepatocyte represents an example of this dilemma. Although the hepatocyte plays a central role in the pathogenesis of many human disorders, the low rate of cell proliferation has limited the use of retrovirus vectors, liposome-DNA complexes, and ligand-based DNA complexes for gene therapy. In models where hepatocyte replication can be enhanced, where the exogenous gene can be transferred to the hepatocyte *in vitro* and the modified cells reinfused, or where transient and/or relatively low level expression of the transferred gene may be sufficient to achieve the therapeutic goal¹⁻³.

To circumvent the requirement for ongoing hepatocyte replication for successful *in vivo* gene transfer, we studied adenovirus (Ad) vectors, which do not require host cell proliferation for expression of the gene to be transferred⁴⁻⁶. Recombinant, replication-deficient Ad vectors can accommodate large exogenous genes and have been used successfully in transferring human genes to the respiratory epithelium *in vivo*, cell targets that also replicate infrequently⁷⁻⁹. Hepatocytes, although rarely, can be targets for clinical adenovirus infection^{10,11}, and because the respiratory epithelium and the liver share the same embryologic origin¹² and genes can be transferred to the neonatal liver by recombinant Ad vectors¹³, we hypothesized that the adult liver could be a target for *in vivo* Ad-mediated gene transfer. From a safety viewpoint, there are extensive epidemiologic data documenting that Ad infections are not associated with human malignancies and live adenovirus vaccines have been used safely in large

human populations for more than two decades¹⁴⁻¹⁶.

In the present study, the possibility of using the Ad to transfer human genes to hepatocytes *in vivo* was evaluated using the *E. coli* lacZ gene and a human α 1-antitrypsin (a1AT) cDNA as model genes. The expression of the lacZ gene, coding for β -galactosidase, serves as a model to identify cells in which there has been successful *in vivo* gene transfer. The expression of the human a1AT cDNA serves as a model of the ability of the Ad vector to confer upon the hepatocyte the capacity to synthesize a typical liver-produced secretory glycoprotein, as directed by the transferred gene. It also provides a convenient protein for evaluation of chronicity of expression by following serum levels of the gene product¹⁷.

Adenoviral transfer to cultured hepatocytes

Primary cultures of rat hepatocytes exposed to the recombinant Ad vectors demonstrated transfer and expression of the exogenous genes. The exposure of cultured hepatocytes to the lacZ containing Ad vector (Ad-RSV β gal; see Methodology) at a multiplicity of infection of 100 pfu per cell for 24 h resulted in expression of β -galactosidase in the majority (62%) of the hepatocytes with no associated change in the morphology of the cells (not shown). Exposure of the cultured hepatocytes to an Ad vector containing the human a1AT cDNA (Ad a1AT)^{18,19} resulted in expression of this cDNA and also caused no morphologic changes (Fig. 1a, b). *In situ* hybridization analysis of uninfected hepatocytes with human a1AT antisense cRNA probe demonstrated no human a1AT mRNA transcripts (background level of grains), that is, the human a1AT antisense probe did no

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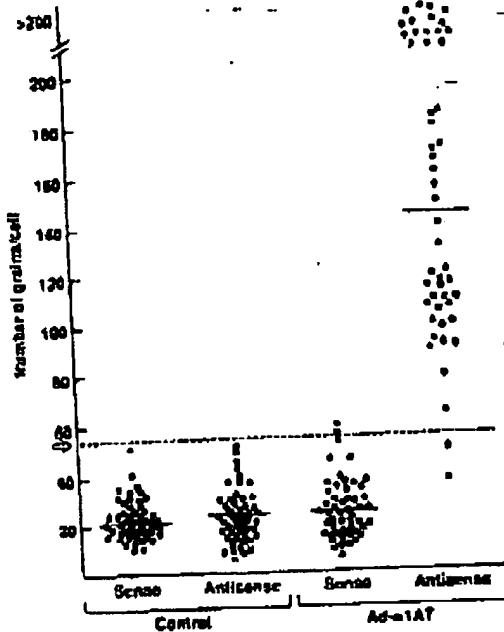
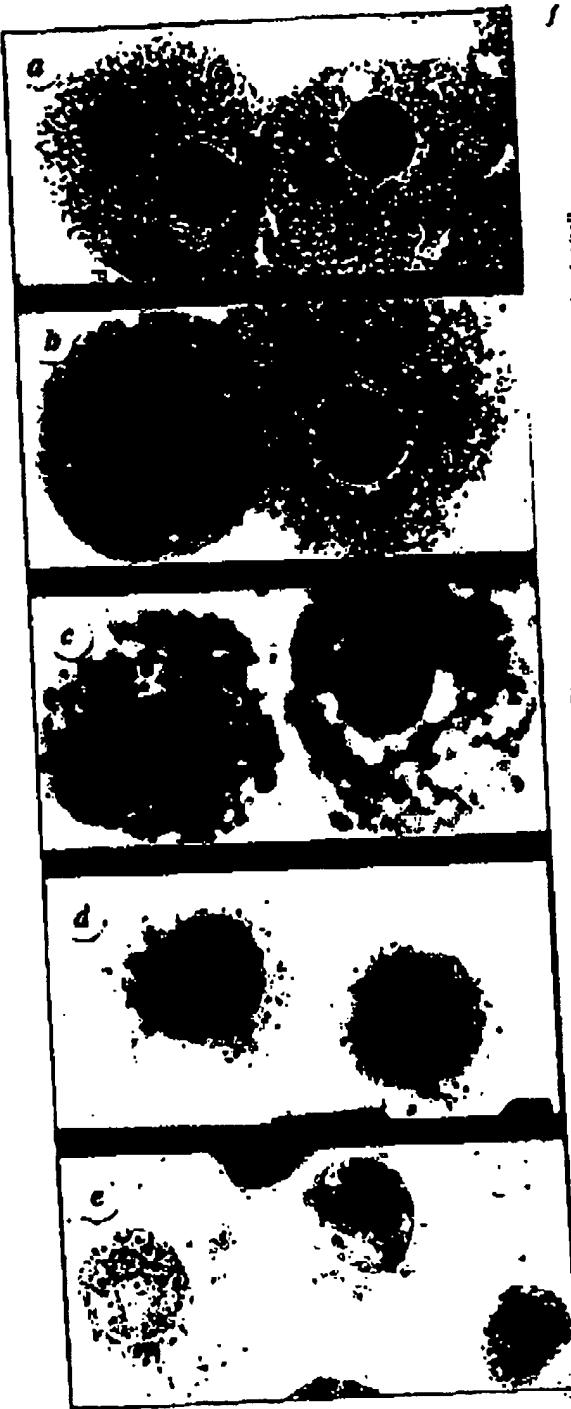


Fig. 1 Expression of human α 1-antitrypsin (α 1AT) mRNA in rat hepatocytes following adenovirus-mediated transfer of the human α 1AT cDNA *in vitro*. Primary cultures of rat hepatocytes were infected *in vitro* with Ad- α 1AT, a replication deficient, recombinant adenovirus (Ad) based on Ad type 5, containing a human α 1AT cDNA driven by the Ad2 major late promoter. *In situ* hybridization evaluation of rat hepatocytes for the presence of human α 1AT mRNA transcripts after 72 h (all original magnifications $\times 1000$). a, Uninfected hepatocytes, Wright-Giemsa. b, Hepatocytes after infection with Ad- α 1AT, Wright-Giemsa. c, Uninfected hepatocytes, [35 S]UTP-labeled antisense human α 1AT cRNA probe. d, Ad- α 1AT infected hepatocytes, antisense probe. e, As d, but evaluated with a control [35 S]UTP-labeled sense α 1AT cRNA probe. f, Quantification of silver grains exposed over control and Ad- α 1AT infected hepatocytes evaluated with the sense and antisense probes. Each symbol represents a randomly selected cell within the defined populations. The arrow and dashed line designate the number of silver grains 3 standard deviations above the mean number of grains exposed over controls; uninfected hepatocytes evaluated with the sense and antisense probes and Ad- α 1AT infected hepatocytes evaluated with the sense probe.

recognize the endogenous rat hepatocyte α 1AT mRNA transcripts (Fig. 1c). Three days after Ad- α 1AT infection, however, the same antisense probe clearly demonstrated human α 1AT mRNA transcripts in rat hepatocytes (increased numbers of grains over the cells, Fig. 1d). The control sense probe demonstrated no transcripts (background level of grains, Fig. 1e). Quantitative assessment of the Ad- α 1AT infected cultures with the

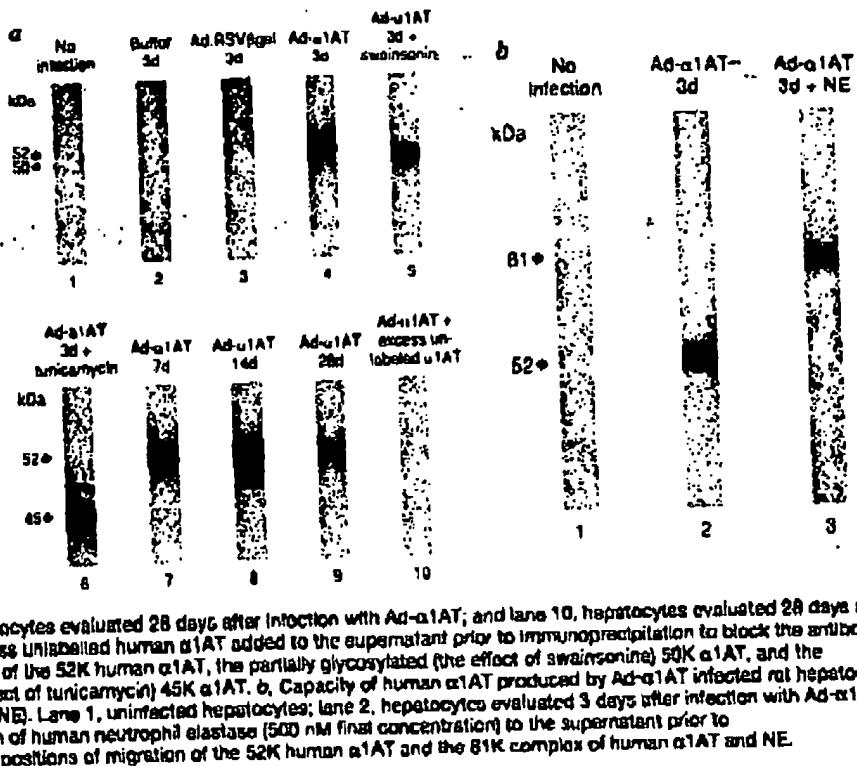
cells had a higher number of autoradiographic grains than parallel cultures evaluated with the sense probe or uninfected hepatocytes evaluated with sense or antisense probes, suggesting that the α 1AT cDNA was transferred to the majority of the cells *in vitro* (Fig. 1f). The viability of Ad- α 1AT infected cultures was similar to uninfected or control Ad vector infected cultures.

Examination of the Ad- α 1AT infected primary rat hepatocytes revealed that they were synthesizing and secreting human α 1AT *de novo* (Fig. 2a), [35 S]methionine labelling for 24 h, and evaluation of the culture supernatants by immunoprecipitation with an anti-human α 1AT antibody demonstrated secretion of human α 1AT with the appropriate migration for mature, glycosylated α 1AT molecules (relative molecular mass 52,000 (M,

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Fig. 2 Evaluation of *in vitro* Ad- α 1AT infected rat hepatocytes for production of human α 1AT and functional analysis of Ad- α 1AT infected rat hepatocyte produced human α 1AT by [35 S]methionine labeling, immunoprecipitation with an anti-human α 1AT antibody, SDS-PAGE and autoradiography. a. Synthesis, secretion and characterization of human α 1AT over time by Ad- α 1AT infected rat hepatocytes. Lane 1, uninfected hepatocytes; lane 2, hepatocytes exposed to viral suspension buffer; lane 3, hepatocytes evaluated 3 days after infection with Ad-RSV β gal; a recombinant, replication deficient Ad containing the *E. coli* lacZ gene; lane 4, hepatocytes evaluated 3 days after infection with Ad- α 1AT; lane 5, as lane 4, but with the addition of swainsonine; lane 6, as lane 4, but with the addition of tunicamycin; lane 7, hepatocytes evaluated 7 days after infection with Ad- α 1AT; lane 8, hepatocytes evaluated 14 days after infection with Ad- α 1AT; lane 9, hepatocytes evaluated 28 days after infection with Ad- α 1AT; and lane 10, hepatocytes evaluated 28 days after infection with Ad- α 1AT but with excess unlabelled human α 1AT added to the supernatant prior to immunoprecipitation to block the antibody. Shown are the positions of migration of the 52K human α 1AT, the partially glycosylated (the effect of swainsonine) 50K α 1AT, and the completely non-glycosylated (the effect of tunicamycin) 45K α 1AT. b. Capacity of human α 1AT produced by Ad- α 1AT infected rat hepatocytes to complex with neutrophil elastase (NE). Lane 1, uninfected hepatocytes; lane 2, hepatocytes evaluated 3 days after infection with Ad- α 1AT; lane 3, as lane 2 but with the addition of human neutrophil elastase (500 nM final concentration) to the supernatant prior to immunoprecipitation. Shown are the positions of migration of the 52K human α 1AT and the 81K complex of human α 1AT and NE.



52K), compare lane 4 to no infection, virus suspension buffer and Ad-RSV β gal infection controls, lanes 1-3). The fact that the secreted α 1AT has the apparent M_r of glycosylated α 1AT is important because glycosylated α 1AT has a half-life in the circulation 50-fold greater than that of non-glycosylated α 1AT²¹. Further evidence that the newly synthesized human α 1AT was glycosylated in a normal fashion came from comparison of α 1AT when the Ad- α 1AT infected hepatocytes were incubated in the presence of swainsonine (an inhibitor of carbohydrate side chain processing in the Golgi) or tunicamycin (an inhibitor of core glycosylation in the rough endoplasmic reticulum); the apparent M_r dropped to 50K (lane 5) and 45K (lane 6), respectively^{21,22}. Ad- α 1AT infected hepatocytes continued to produce α 1AT for at least 28 days (lanes 4, 7-9). The specificity of the antibody for human α 1AT was demonstrated by adding excess unlabelled human α 1AT to the immunoprecipitation reaction (lane 10). When neutrophil elastase (NE, the natural substrate of α 1AT) was added to the supernatants of Ad- α 1AT infected hepatocytes, the expected 81K α 1AT-NE complex²³ were observed (Fig. 2b).

Adenoviral transfer *in vivo*

In vivo gene transfer to the liver was first evaluated by introduction of Ad-RSV β gal or controls (virus suspension buffer, or control viruses Ad-dl312, Ad- α 1AT, or Ad-CFTR (see Methodology)) into the liver via intraportal injection (Fig. 3). Rats observed for up to 90 days showed no adverse effects (altered survival, recovery time post-surgery, coat appearance or general activity level) compared to controls. Livers exposed to the Ad vectors *in vivo* revealed no gross or microscopic anatomical derangements when compared to the livers recovered

from non-injected or virus suspension buffer injected animals. Livers recovered 3 days after infusion of an equivalent volume of virus suspension buffer or 10¹⁰ pfu Ad- α 1AT revealed no evidence of lacZ gene expression (Fig. 3a, b). Likewise, infusion of 10¹⁰ pfu of Ad-dl312 or Ad-CFTR or no infusion at all resulted in no β -galactosidase activity (not shown).

In contrast, livers recovered from rats having received 10¹⁰ pfu intraportal Ad-RSV β gal 3 days previously showed expression of the exogenous gene in hepatocytes, as seen by the nuclear blue staining, characteristic of β -galactosidase activity as directed by the lacZ gene together with the SV40 nuclear localization signal. The distribution of positive cells was discontinuous: some areas showed randomly distributed single positive cells (Fig. 3c), other areas showed more closely spaced (Fig. 3d-f) or clumped (Fig. 3f, g) of blue staining hepatocytes. Some specimens had broad areas with no positive cells. Interestingly, Kupffer cells, biliary tract cells or hepatic vascular endothelial cells expressed β -galactosidase activity. Examination of blindly selected samples of liver exposed to intraportal Ad-RSV β gal documented an average of \pm 0.2% (mean \pm SEM; 661 of 59,819 total hepatocytes scored in a total of 2 livers) of hepatocytes expressing lacZ gene. Animals receiving intraportal Ad-RSV β gal showed no evidence of β -galactosidase activity in the lung, brain, kidney, spleen, muscle or testis (not shown). Endogenous thyroid cytoplasmic β -galactosidase activity was uniformly present in Ad-RSV β gal and control animals and served as a positive control²⁴.

In contrast to direct intraportal administration of Ad-RSV β gal vector, attempts at *in vivo* adenoviral transfer to the liver by intravenous injection of 10¹⁰ Ad-RSV β gal ($n = 5$) or, as control, virus suspension



Fig. 3 *In vivo* transfer of the *E. coli* lacZ gene to rat hepatocytes. Three days after Ad.RSVβgal was infused into the portal vein, the liver was evaluated for the presence (blue staining) of the lacZ product β-galactosidase using X-gal stain. a, Liver section from a rat not receiving an intraportal infusion, X-gal stain, haematoxylin and eosin, $× 200$. b, Liver section 3 days after intraportal infusion of Ad-α1AT, X-gal, haematoxylin and eosin $× 400$. c, Liver section 3 days after intraportal infusion of Ad.RSVβgal, X-gal, haematoxylin and eosin, $× 200$. d, and e, As c but $× 400$. f, Liver section 3 days after intraportal infusion of Ad.RSVβgal, X-gal and eosin, $× 1,000$. g, As c, but $× 1,000$.

or 10^6 pfu of Ad-dl312, Ad.RSVβgal, or Ad-CFTR), hepatocytes were isolated at intervals 3–90 days thereafter and the production of α1AT analysed by [35 S]methionine labelling and immunoprecipitation. The de novo synthesis and secretion of human α1AT protein was documented by the immunoprecipitation of the expected 52K human [35 S]-labelled α1AT from cultures of rat hepatocytes recovered by collagenase perfusion at 3, 7, 14, 28, 60 and 90 days after intraportal infusion of Ad-α1AT (not shown). No human α1AT was immunoprecipitated from similar hepatocyte supernatants from uninfected rats or other controls (other Ad vectors or viral suspension buffer). The immunoprecipitable signal eliminated by excess unlabelled human α1AT added to the immunoprecipitation reaction, confirmed that this was indeed human α1AT.

Expression of the *in vivo* transferred human α1AT cDNA was documented *in vivo* by quantification of serum levels of human α1AT with a human α1AT-specific ELISA (Fig. 4). Human α1AT in the circulation of rats having received intraportal Ad-α1AT averaged $215\text{--}380\text{ ng ml}^{-1}$ for up to 28 days ($n = 25$, $p < 0.005$ compared with controls over the same time period using the two-tailed Student's *t*-test; the normal level of endogenous rat α1AT in the circulation is 1.1 mg ml^{-1} (ref. 38)). A few 'anecdotal' animals demonstrated detectable human α1AT in the circulation for as long as 60 or 90 days. No human α1AT was found in serum samples from control rats ($n = 47$), including those receiving no intraportal infusion and those infused with control Ad vectors or viral suspension buffer. Survival was identical in all injected animal groups: sham, viral suspension buffer, control adenoviruses and Ad-α1AT.

Discussion

It is clear that replication-deficient Ad vectors can be used to transfer exogenous genes to hepatocytes *in vivo*, including genes coding for an intracellular functional protein (*E. coli*-galactosidase) and a glycosylated, secreted functional protein (human α1AT). Our data support a new strategy for transfer and expression of exogenous genes into adult hepatocytes *in vivo* without prior perturbation of the predominant G₀ cell cycle state of the adult hepatocyte. This obviates the use of stimulants for hepatocytes such as hepatotoxins or resections, which have significant drawbacks for adaptation to human gene transfer. Several important topics are relevant to the potential usefulness of an Ad vector system for *in vivo* human gene transfer to the liver, including: 1) organ specificity; 2) extent of expression; 3) chronicity of expression and 4) safety.

($n = 3$) into the tail veins of adult rats were unsuccessful. Livers recovered 3 days after injection from either group did not show β-galactosidase activity (not shown).

As another approach to assessing the *in vivo* transfer of the lacZ gene to hepatocytes, we recovered hepatocytes by collagenase perfusion¹ of the liver 3 days after intraportal injection of Ad.RSVβgal. Staining with X-gal revealed a population of hepatocytes with nuclear blue staining consistent with the *in vivo* transfer of the lacZ gene (not shown). Aside from the expression of β-galactosidase activity, this population was in no other way distinguishable from hepatocytes recovered by the same methodology from uninfected rats or rats injected with Ad-dl312, Ad-CFTR, Ad-dl312 or buffer, which stained negative.

In vivo transfer of α1-antitrypsin

Consistent with the observations obtained with Ad.RSVβgal, *in vivo* intraportal infusion of Ad-α1AT produced synthesis of the human α1AT in hepatocytes and the secretion of human α1AT into the circulation. No gross or microscopic anatomical rearrangement of these livers was found compared to controls. Following intraportal administration of Ad-α1AT (10^6 pfu) or controls (an equivalent volume viral suspension buffer,

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Organ specificity to the liver was achieved by local administration. The Ad enters cells by interacting with cell surface receptors that recognize the fibre-penton structure of the virus capsid¹³⁻¹⁷. Following internalization, the virus is released from the virus receptor endosome and the virus moves to the nucleus¹⁸⁻²¹. The specificity for cell targets is likely a function of receptor density, Ad type and local immune surveillance. For targeting to the liver, the most direct approach is to infuse the Ad vector into the hepatic vascular supply such as the portal vein as used here. Importantly, this approach successfully targeted hepatocytes, as no Kupffer cells, biliary tract cells or hepatic vascular endothelial cells expressed the *lacZ* gene following infusion of Ad.RSV β gal. Moreover, the expression was liver-specific.

Adenovirus-mediated gene transfer for amelioration of hepatic enzyme deficiency has been attempted in other model systems. In a mouse model of ornithine transcarbamylase deficiency (OTC) (a gene normally expressed at a relatively low level in liver), systemic administration of an adenovirus vector containing a rat OTC cDNA to newborn mice resulted in some animals with enhanced levels of hepatic enzyme activity and some exhibiting phenotypic correction¹¹. Systemic administration of adenovirus vectors to newborn mice, however, results in the exogenous gene expression in multiple organs including lung, liver, intestine, heart and skeletal muscle¹⁸⁻²². Moreover, in transgenic mouse studies where OTC transgenes were expressed predominantly in the intestine, the phenotypic defect was also corrected^{14,16} raising the question of the contribution of enzyme activity from sites other than the liver. In the current study, adult rats were given an Ad vector directly to the liver via intraportal injection and expression was limited to the liver.

The relatively low levels of α 1AT achieved (Fig. 4) in comparison with a level that would be required for therapy (>0.8 mg ml⁻¹) must be considered with the knowledge that, unlike OTC, the α 1AT gene is expressed in hepatocytes at a high level. Two other fundamental differences exist between the current study and the newborn mouse model. First, adult rats are not necessarily comparable to newborn mice in terms of susceptibility to *in vivo* gene transfer. Second, we determined the serum concentration of α 1AT whereas in the newborn mouse model only the activity of the (intra-mitochondrial) OTC was assessed. Despite these differences, a comparison of the studies suggests that direct intraportal administration to adult yields local expression at a level less than that of a highly expressed protein (α 1AT), whereas systemic administration to neonates results in expression in multiple organs of sufficient activity to produce phenotypic correction for a relatively low level expressed protein (OTC). These concepts will have to be taken into account in designing gene transfer strategies to specific organs.

The extent of expression of the *lacZ* gene within the liver following intraportal Ad.RSV β gal infusion was patchy, with approximately 1% of hepatocytes demonstrating β -galactosidase activity 3 days after infusion. However, >60% of hepatocytes in primary cultures expressed β -galactosidase when infected *in vitro* at a multiplicity of infection of 100 pfu per cell with the same vector, suggesting that the extent of expression *in vivo* may be determined by the amount of functional

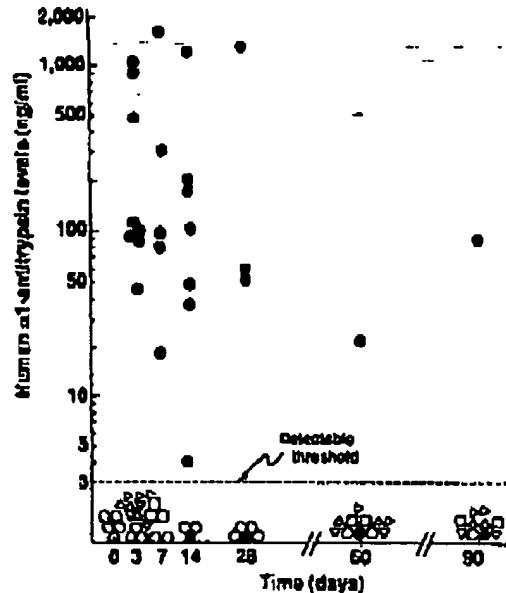


Fig. 4 Levels of human α 1AT in the circulation of rats after intraportal administration of Ad- α 1AT. Ad- α 1AT (10^8 pfu) or Ad control vectors (10^8 pfu each) or buffer was infused into the portal vein of anesthetized rats. Each symbol represents the mean of quadruplicate determinations from an individual rat. Shown are data for Ad- α 1AT infected animals (○), and a variety of controls: (□), rats receiving no intraportal infusion; (▽), rats receiving intraportal virus suspension buffer; (△), Ad-d312; (▲), Ad-CFTR; (◇), Ad.RSV β gal. No α 1AT was detected in the preparations of Ad- α 1AT used for infection.

vector reaching each hepatocyte. Since the rat liver contains approximately 10^9 hepatocytes²³, infusion of 10^8 pfu will permit, at most, 10 pfu per cell. Moreover, "pfu" for the Ad vectors are defined on a cell line (293, transformed human embryonal kidney cells)²⁴ which provides E1 function in trans to the replication-deficient Ad vectors. This *in vitro* titration method may not reflect the infectivity of rat hepatocytes *in vivo*. The patchy *in vivo* expression may also be affected by the distribution of the perfuse bolus after injection into the portal vein. Because the intraportal adenovirus strategy does not require prior hepatic damage to enhance hepatocyte replication^{11,12,14,15} or partial hepatectomy to harvest hepatocytes for *in vitro* gene transfer and reimplantation^{18,25}, it avoids the natural morbidity and mortality of these procedures. Furthermore, optimization of the *in vivo* adenovirus delivery system (for example, by higher dosages or repeated administration) may offer significant increases in exogenous gene expression and/or the duration of expression of the transferred gene.

The expression achievable using the present strategy would not be sufficient to treat human α 1AT deficiency but might possibly be sufficient to provide enough recombinant molecules (alone or with conventional therapy) to treat a variety of genetic disorders, including the factor VIII and factor IX haemophilias, deficiency disorders of the urea cycle, disorders of amino acid metabolism, carbohydrate metabolism, or porphyria related disorders^{1,17-20}. The extent of expression seen here

may allow therapy for disorders such as LDL receptor mutations, should the receptor systems be sufficiently potent^{12,13,17-19}, or in females with X-linked disorders where the pattern of X-inactivation has left them with suboptimal levels of gene expression¹⁷⁻¹⁹.

In reference to chronicity of expression, the available evidence suggests that the Ad genes likely function in an extra-chromosomal fashion — little is integrated in the genome²⁰. While this has advantages as a vector system in minimizing insertional mutagenesis, the chronicity of infection may be limited by the rate of cell turnover. The expression for at least 4 weeks of the human α 1AT cDNA following *in vivo* transfer to the liver is consistent with the slow turnover of adult hepatocytes *in vivo*²¹. Alternatively, the exogenous gene may persist in the hepatocyte but expression may decline as has been observed for genes transferred to cells *in vitro* by recombinant retrovirus vectors and then reimplanted²². This may be obviated, however, by repeated dosages.

Finally, the safety of replication deficient recombinant adenovirus vectors *in vivo* in humans is unknown, and further study will be needed to resolve this issue. There is evidence to suggest however, that safe use may be possible. At no time interval after *in vivo* transfer with Ad vectors was there gross or microscopic evidence of hepatic damage, regenerative or degenerative changes, inflammatory responses, or evidence of neoplastic changes. Importantly, human Ad infection is not associated with malignancy. The Ad vectors are constructed to be replication deficient; while it is conceivable that exogenous wild type Ad could complement the deleted region to propagate the recombinant virus, Ad hepatic infection is a rare clinical event in humans^{23,24}, suggesting this may not be an important safety issue. Finally, oral Ad vaccines have been safely used in humans for decades with an excellent safety record²⁵.

Methodology

Hepatocyte culture. Hepatocytes were isolated from Sprague Dawley rats (250–350 g) by hepatic perfusion with 140 U ml^{-1} collagenase (Type IV, Sigma), 10 mM HEPES, pH 7.4 in Williams' media E (GIBCO)²⁶. Recovered cells were allowed to adhere for 4 h on tissue culture plates (Becton Dickinson, Lincoln Park, New Jersey). In Dulbecco's modified Eagle medium (DMEM) mixed 1:1 with Waymouth's medium (both from GIBCO) including dexamethasone 20 $\mu\text{g ml}^{-1}$ (Sigma), insulin 5 $\mu\text{g ml}^{-1}$, transferrin 5 $\mu\text{g ml}^{-1}$, selenite acid 5 mg ml^{-1} (ITS, Collaborative Research, Bedford, Massachusetts), gentamicin 10 $\mu\text{g ml}^{-1}$, 10 mM HEPES pH 7.4, and 10% heat inactivated fetal calf serum (Hyclone Laboratories, Logan, Utah). Nonadherent cells were removed by washing twice with $\text{Ca}^{2+}/\text{Mg}^{2+}$ -free Hank's balanced salt solution (HBSS). Histologic evaluation demonstrated >98% of the cells had the typical appearance of hepatocytes. Cultures were continued in maintenance media that differed from plating media by the deletion of fetal calf serum and the inclusion of bovine serum albumin 1.25 mg ml^{-1} and linoleic acid 5.35 $\mu\text{g ml}^{-1}$ (ITS+, Collaborative Research).

Adenovirus vectors. Adenoviruses were prepared, purified and titered as described^{27,28}. Ad- α 1AT is an Ad5 E1a, E3 deletion based vector with the Ad2 major late promoter and the human α 1AT

cDNA²⁹. Ad-RSV β gal is an Ad5 E1, E3 deleted replication deficient, recombinant adenovirus identical to Ad-CFTR (see ref. 25) except that in place of the Ad5 major late promoter and human cystic fibrosis transmembrane conductance regulator (CFTR) cDNA there is the Rous sarcoma virus long terminal repeat, the SV40 nuclear localization signal, and the *E. coli* lacZ gene^{26,28}. Ad-dl312 is an Ad5 E1a deletion mutant without an exogenous gene²⁷. Ad-CFTR is an Ad5 based, E1, E3 deletion vector otherwise similar to Ad- α 1AT, but with the human (CFTR) cDNA²⁸ instead of the α 1AT cDNA.

In vivo gene transfer to hepatocytes. After die change to maintenance media, hepatocyte cultures were infected with Ad- α 1AT or Ad-RSV β gal at a multiplicity of infection of 100 pfu per cell. Control, replicate cultures were exposed to equivalent volumes of virus suspension buffer (10 mM Tris-HCl, 1 mM MgCl₂, 10% glycerol, pH 7.4) or media alone. For the *in situ* analyses hepatocytes were recovered after 72 h by exposure to trypsin-ethylene diamine tetraacetic acid (2 min, 37 °C; Trypsin/Versene, Biofluids, Rockville, Maryland). gentle scraping and cytocentrifugation. Human α 1AT mRNA transcripts were evaluated in cytocentrifuge preparations with [³⁵S]UTP-labelled sense and antisense cDNA probes of similar specific activity and concentration (1.2×10^5 dpm ml^{-1}) prepared from a human α 1AT cDNA insert in pGEM-3Z (Promega). After hybridization, preparations were exposed to autoradiography film for 4 days and counterstained with Wright-Giemsa^{26,28}. For evaluation of human α 1AT synthesis, secretion and function the hepatocytes were isolated and infected as above. After 3, 7, 14, or 28 days the cells were incubated (24 h, 37 °C) with [³⁵S]methionine (250 $\mu\text{Ci ml}^{-1}$ (>1000 Ci mmol⁻¹; New England Nuclear)). To demonstrate that the human α 1AT was glycosylated in a normal fashion, parallel cultures of Ad- α 1AT infected hepatocytes were labelled with [³⁵S]methionine after the addition of tunicamycin (10 $\mu\text{g ml}^{-1}$, an inhibitor of core glycosylation) or swainsonine (5 $\mu\text{g ml}^{-1}$, an inhibitor of distal glycosylation processing (both from Boehringer Mannheim Biochemicals)). Supernatants were evaluated by immunoprecipitation with a rabbit anti-human α 1AT antibody (Boehringer Mannheim). SDS-polyacrylamide gel electrophoresis and autoradiography. The function of the human α 1AT produced was demonstrated by addition of human neutrophil elastase (Elastis Products, Owensville, Maryland) prior to immunoprecipitation²⁸.

In vivo gene transfer to liver. Sprague Dawley rats (250–350 g) were anaesthetized with a mixture of ketamine HCl and xylazine. The peritoneal cavity was incised and the portal vein isolated and injected with 10⁶ pfu of Ad-RSV β gal or Ad- α 1AT. The abdomen was closed and the rats allowed to recover. After 3 days, some animals were sacrificed, bled, and the liver fixed *in situ* by perfusion with PBS (4 °C) followed by 4% paraformaldehyde (Fisher Scientific, Fair Lawn, New Jersey) in PBS (4 °C). Livers were then minced into 2 mm fragments, and evaluated for β -galactosidase activity by incubation in X-gal staining solution (5 mM K₄Fe(CN)₆, 5 mM K₃Fe(CN)₆, (both from Sigma), 5 mM MgCl₂ (Mallinckrodt), and 200 $\mu\text{g ml}^{-1}$ X-gal (5-bromo-4-chloro-3-indolyl- β -D-galactopyranoside, Boehringer Mannheim))²⁸. After staining, tissues were embedded in paraffin, sectioned, adhered to glass slides and counterstained with hematoxylin and eosin or eosin alone. Controls included no injections or intraperitoneal injection of Ad-dl312, Ad-CFTR or an equivalent volume of virus suspension buffer. The remaining control animals and those receiving intraperitoneal Ad- α 1AT or Ad-RSV β gal were sacrificed at 3, 7, 14, 28, 60 and 90 days after injection, bled, hepatocytes isolated (described above), and the *de novo* synthesis and secretion of human α 1AT analyzed by [³⁵S]methionine labelling and immunoprecipitation²⁸. The *in vivo* expression of the Ad- α 1AT transferred α 1AT cDNA was analyzed in the serum samples with a human α 1AT-specific ELISA with a sensitivity of ≥ 3 ng ml^{-1} .

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